



Graft-Versus-Host Disease

Mechanism of Disease

Presentation Overview

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GVHD Overview



GVHD Results From Immunologic Attack on Recipients' Target Organs or Tissues by Donor Allogeneic T Cells¹

aGVHD

- Activated donor T cells damage host epithelial cells and mucous membranes after an increase in inflammation following the conditioning regimen for HSCT^{2,3}
- Organs affected: primarily skin, gut, and liver³

cGVHD

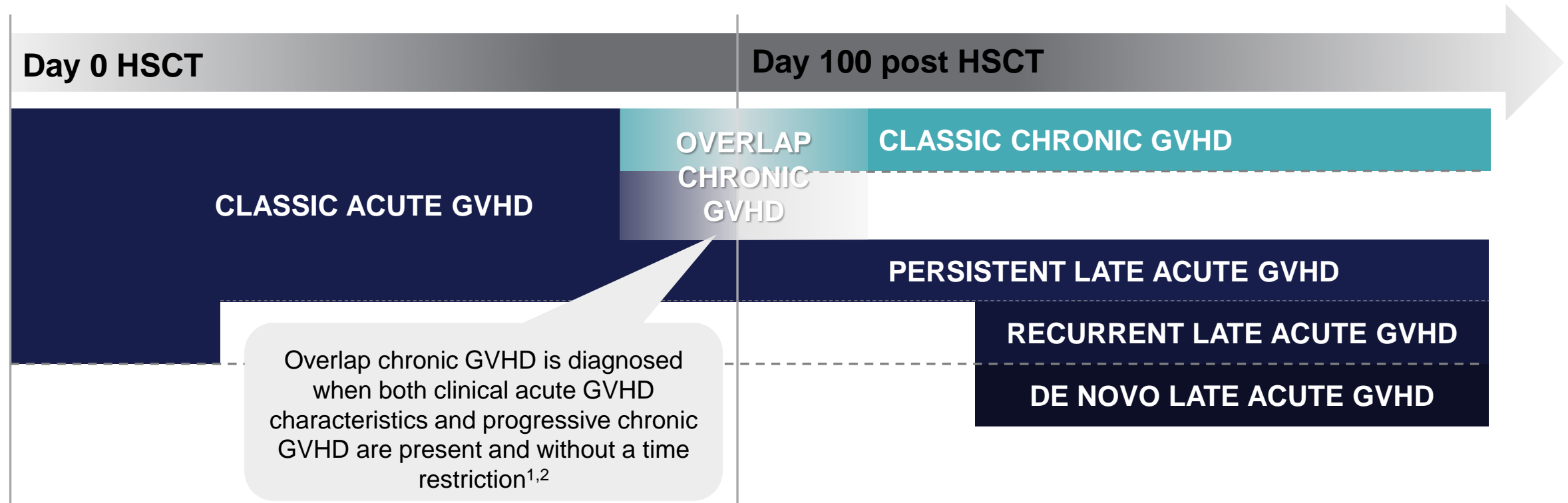
- Complex immune-mediated pathology of cGVHD involves T cells, B cells, macrophages, and fibroblasts^{4,5}
- Displays more autoimmune and fibrotic features than aGVHD^{4,5}
- Organs affected: primarily oral and ocular mucosal surfaces; may also affect the skin, gut, liver, lungs, and kidneys^{2,5}

- Loss of central tolerance, autoantibody production, and fibrosis in cGVHD are thought to distinguish cGVHD from aGVHD^{5,6}
- “Overlap cGVHD” is characterized by clinical features of both aGVHD and cGVHD^{5,7}

aGVHD, acute graft-versus-host disease; cGVHD, chronic graft-versus-host disease; GVHD, graft-versus-host disease; HSCT, hematopoietic stem cell transplantation.

1. Blazar BR, et al. *Nat Rev Immunol*. 2012;12:443-458. 2. Jacobsohn DA, Vogelsang GB. *Orphanet J Rare Dis*. 2007;2:35. 3. Vogelsang GB, et al. *Annu Rev Med*. 2003;54:29-52. 4. McDonald-Hyman C, et al. *Sci Transl Med*. 2015;7:280rv2. 5. Cooke KR, et al. *Biol Blood Marrow Transplant*. 2017;23:211-234. 6. Abboud R, et al. *Ther Adv Hematol*. 2020;11:1-13. 7. Lee SJ. *Blood*. 2017;129:30-37.

Classification of GVHD



Acute and chronic GVHD are considered distinct clinical syndromes without a time restriction¹

Figure adapted from Lee SJ. *Blood*. 2017;129:30-37.¹

Note: Box sizes do not reflect relative prevalence or incidence.

1. Lee SJ. *Blood*. 2017;129:30-37. 2. Filipovich AH, et al. *Biol Blood Marrow Transplant*. 2005;11:945-956.

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Acute GVHD

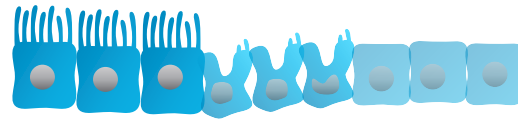
Mechanism of Disease



Clinical Manifestations of aGVHD

- In aGVHD, the conditioning regimen causes tissue damage, which activates T cells¹
- Activated T cells expand and differentiate into cytotoxic effector T cells¹
- Effector T cells migrate from lymphoid tissues to cause organ damage¹

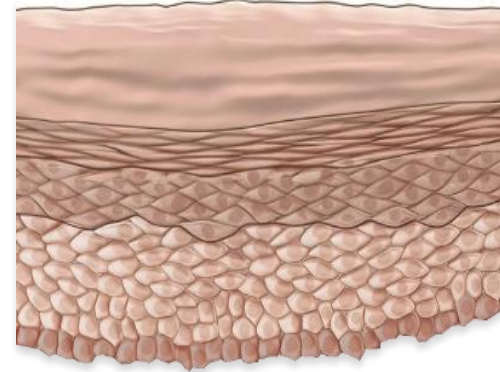
Gut



Colonic mucosa may develop withered and necrotic crypts. Large deep ulcers are seen with mucosal sloughing and loss of epithelium²

Diarrhea

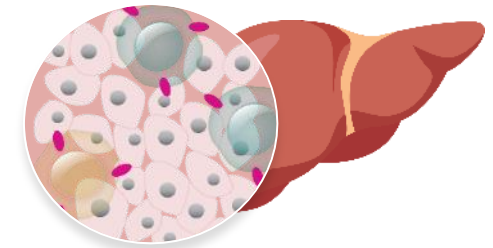
Skin



Cutaneous manifestations in aGVHD can affect the skin causing erythematous maculopapular rashes on the face, ears, palms and soles¹

Rash

Liver



Bile duct damage may occur in hepatic GVHD. The epithelial cells of the bile duct may show eosinophilic cytoplasm and variable nuclear hyperchromasia with crowding³

Hyperbilirubinemia

Most common symptoms⁴

1. Rodrigues KS, et al. *Am J Clin Dermatol*. 2018;e19:33-50. 2. Naymagon S, et al. *Nat Rev Gastroenterol Hepatol*. 2017;14:711-726. 3. Stueck AE, et al. *Mod Pathol*. 2018;31:442-451. 4. Ferrara JLM, et al. *Lancet*. 2009;373:1550-1561.

aGVHD: Epidemiology and Significance

Serious complications of allo-HSCT with significant morbidity and mortality^{1,2}

- Incidence of acute GVHD varies by donor type and prophylaxis regimen
 - MSD: approximately 30% to 40%³
 - MUD or mismatched relative: approximately 20% to 50%⁴⁻⁵
 - Standard prophylaxis (CNI+MTX/MMF±ATG): ranges from 30% to 60%^{6,7}
- In recent studies, death due to complications of aGVHD was reported in 16% to >30% of patients⁸⁻¹⁰

Immunosuppression with corticosteroids is first-line therapy but is insufficient for most patients^{11,12}

- Less than 50% of patients with aGVHD experience clinically relevant responses to first-line therapy¹¹

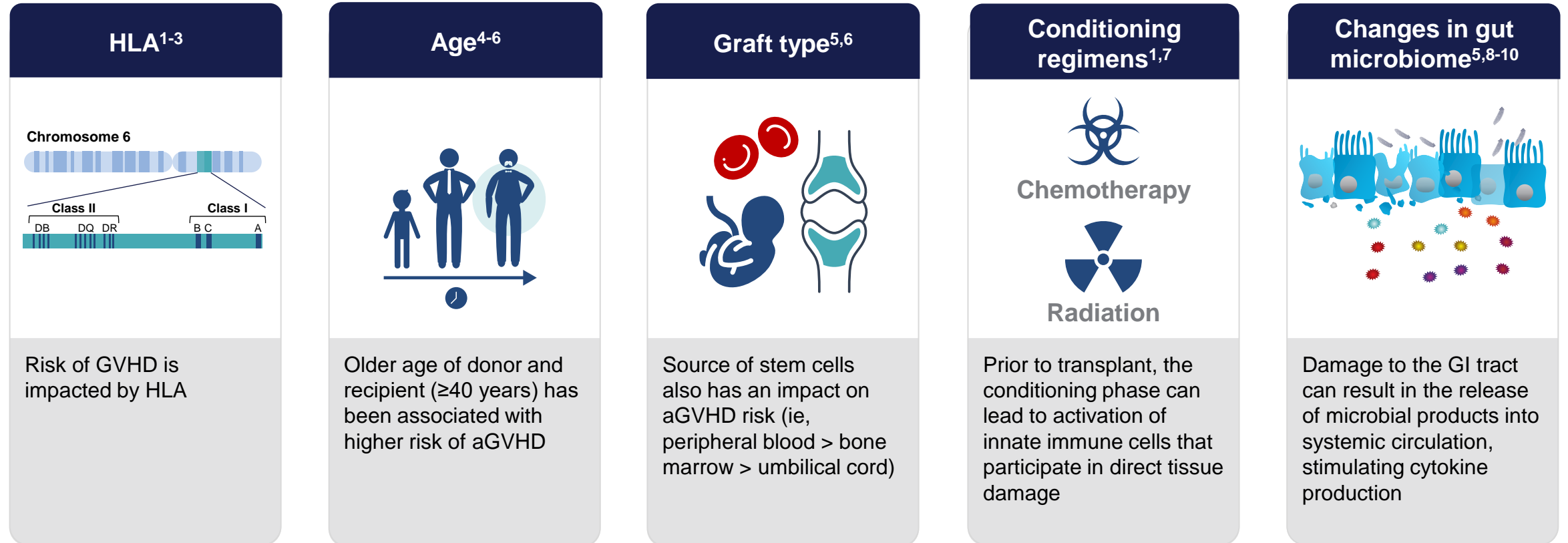
May rapidly progress to steroid-refractory disease¹³

- Steroid-refractory aGVHD has been reported to have a mortality rate of approximately 35%⁹

allo-HSCT, allogeneic hematopoietic stem cell transplantation; ATG, antithymocyte globulin; CNI, calcineurin inhibitor; MMF, mycophenolate mofetil; MSD, matched sibling donor; MTX, methotrexate; MUD, matched unrelated donor.

1. Jagasia M, et al. *Blood*. 2012;119:296-307. 2. Jacobsohn DA, Vogelsang GB. *Orphanet J Rare Dis*. 2007;2:35. 3. Nagler A, et al. *Transplant Cell Ther*. 2022;28(2):86.e1-86.e8. 4. Nagler A, et al. *Clin Cancer Res*. 2021;27(3):843-851. 5. Luznik L, et al. *J Clin Oncol*. 2022;40(4):356-368. 6. Jamy O, et al. *Blood*. 2023;142(12):1037-1046. 7. Malard F, et al. *Nat Rev Dis Primers*. 2023;9(1):27. 7. Yu J, et al. *Curr Med Res Opin*. 2019;35:983-988. 8. Yu J, et al. *Biol Blood Marrow Transplant*. 2020;26:600-605. 9. Ramdial JL, et al. *Bone Marrow Transplant*. 2021;56:2005-2012. 10. Garnett C, et al. *Ther Adv Hematol*. 2013;4:366-378. 11. Magenau J, et al. *Br J Haematol*. 2016;173:190-205. 12. Schoemans HM, et al. *Bone Marrow Transplant*. 2018;53:1401-1415.

Risk Factors for Development of aGVHD

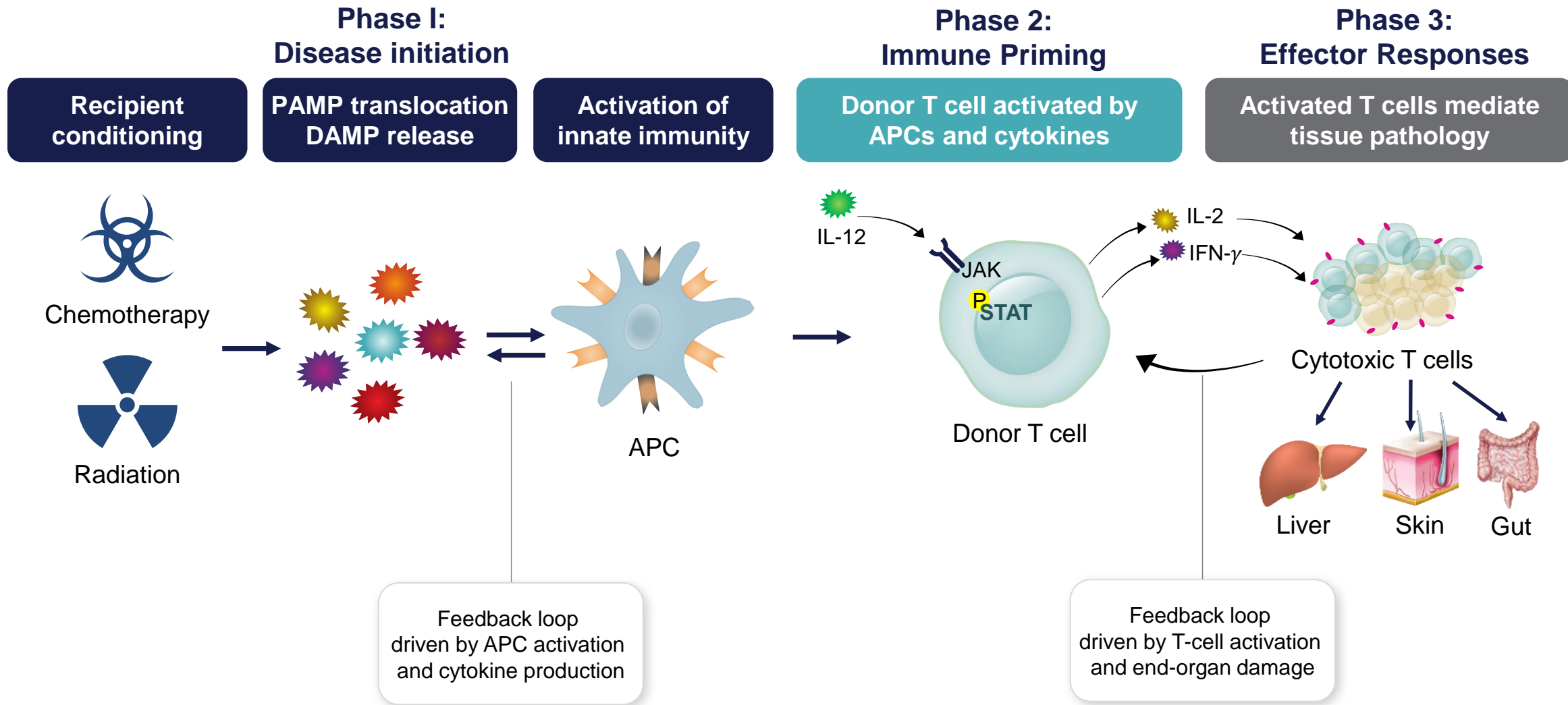


GI, gastrointestinal; HLA, human leukocyte antigen.

1. Ferrara JLM, et al. *Lancet*. 2009;373:1550-1561. 2. Loiseau P, et al. *Biol Blood Marrow Transplant*. 2007;13:965-974. 3. Flowers MED, et al. *Blood*. 2011;117:3214-3219. 4. Nash RA, et al. *Blood*. 1992;80:1838-1845. 5. Nasserredine S, et al. *Anticancer Res*. 2017;37:1547-1555. 6. Cutler C. *Hematology*. 2008;3(part 1):1-12. 7. Blazar BR, et al. *Nat Rev Immunol*. 2012;12:443-458. 8. Whangbo J, et al. *Bone Marrow Transplant*. 2017;52:183-190. 9. Taur Y, et al. *Blood*. 2014;124:1174-1182. 10. Jenq RR, et al. *J Exp Med*. 2012;209:903-911.

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Overview of aGVHD Pathophysiology



APC, antigen presenting cell; DAMP, damage-associated molecular pattern; IFN, interferon; IL, interleukin; JAK, Janus kinase; P, phosphorylated; PAMP, pathogen-associated molecular pattern; STAT, signal transducer and activator of transcription.

Hill GR, et al. *Ann Rev Immunol.* 2021;39:19-49.

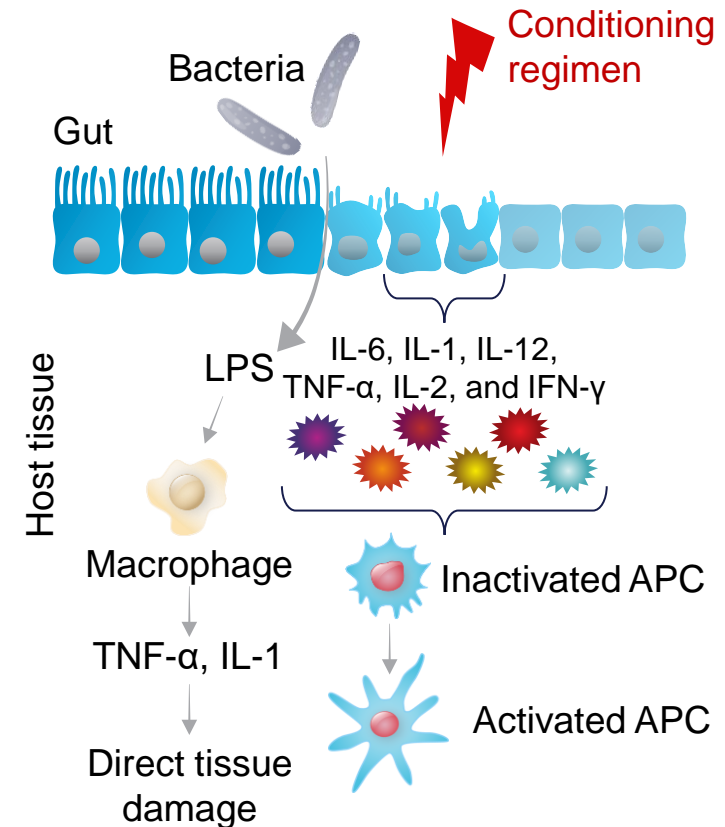


Disease Initiation: Damage to Host Tissue Activates Donor and Host APCs

- Occurs due to HLA mismatch, conditioning, and/or other factors^{1,2}
- Conditioning enables host recipients to receive immunocompetent T cells^{3,4}
- Conditioning-mediated tissue injury to host mucosa, skin, and liver leads to cytokine release and innate immune activation, initiating GVHD^{3,5-8}
- Activated donor and host APCs then release their own cytokines, resulting in a positive feedback loop^{2,4}

Disease initiation caused by recipient conditioning and other factors^{1,8-10}

Allo-antigens are produced

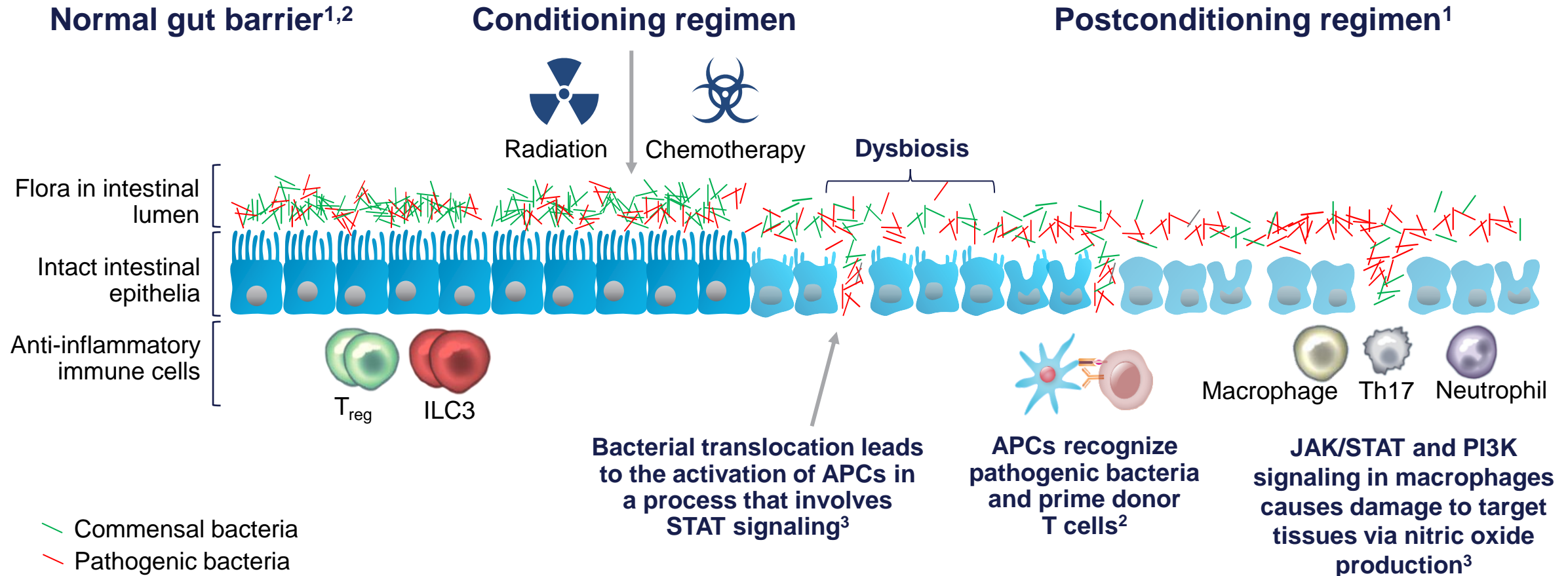


LPS, lipopolysaccharide; TNF, tumor necrosis factor.

1. Nassereddine S, et al. *Anticancer Res.* 2017;37:1547-1555. 2. McDonald-Hyman C, et al. *Sci Transl Med.* 2015;7:280-282. 3. Ferrara JL, et al. *Stem Cells.* 1996;14:473-489. 4. Blazar BR, et al. *Nat Rev Immunol.* 2012;12:443-458. 5. Toubai T, et al. *Front Immunol.* 2016;7:539. 6. Markey KA, et al. *Blood.* 2014;124:354-362. 7. Zeiser R, Blazar BR. *N Engl J Med.* 2017;377:2167-2179. 8. Abboud R, et al. *Ther Adv Hematol.* 2020;11:1-13. 9. Schroeder MA, DiPersio JF. *Dis Model Mech.* 2011;4:318-333. 10. Ferrara JLM, et al. *Lancet.* 2009;373:1550.

Disease Initiation: Changes in Gut Microbiome Contribute to aGVHD

1

Disease
Initiation

ILC, innate lymphoid cell; PI3K, phosphoinositide 3 kinase; Th, T helper; T_{reg}, regulatory T cell.

1. Rafei H, Jenq RR. *Blood*. 2020;136:401-409. 2. Chen Y, et al. *J Immunol Res*. 2015;2015:145859. 3. Abboud R, et al. *Ther Adv Hematol*. 2020;11:2040620720914489.

Donor T-Cell Activation and Expansion

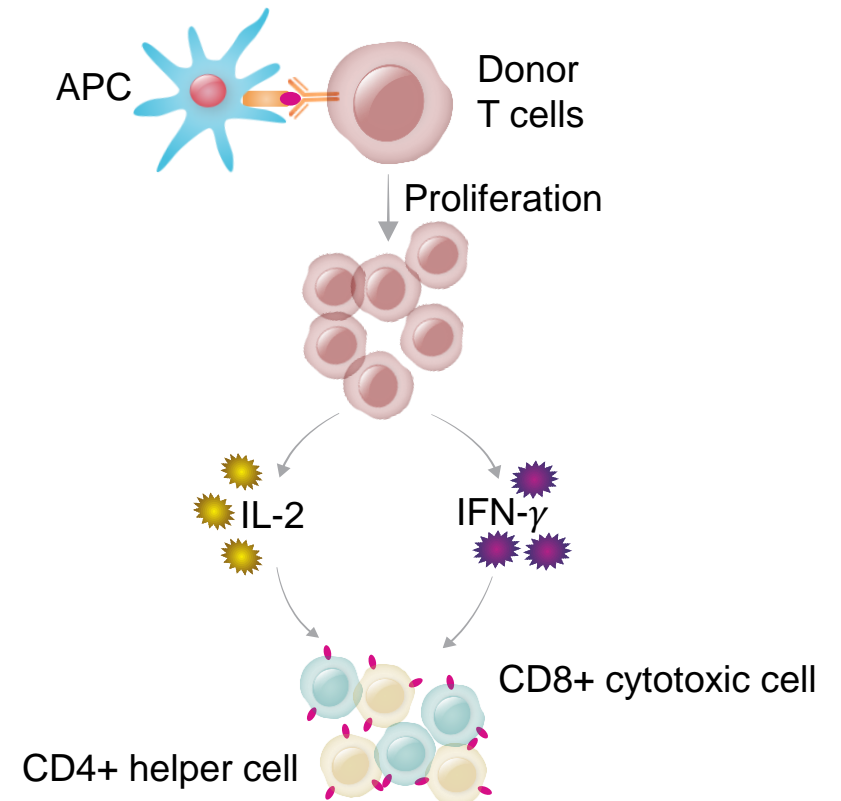
2

Immune
priming

- T-cell receptors and co-stimulatory molecules (eg, CD28) on donor T cells interact with ligands on the surface of the activated APCs, leading to donor T-cell activation¹
- Inflammatory cytokines produced in response to these activation signals subsequently stimulate T-cell expansion and differentiation^{1,2}
- Signaling through the IFN- γ receptor and the JAK/STAT pathway results in increased T-cell trafficking to the gut, liver, and skin²

Donor T-cell activation²⁻⁴

Production of cytolytic donor effector T cells



CD, cluster of differentiation.

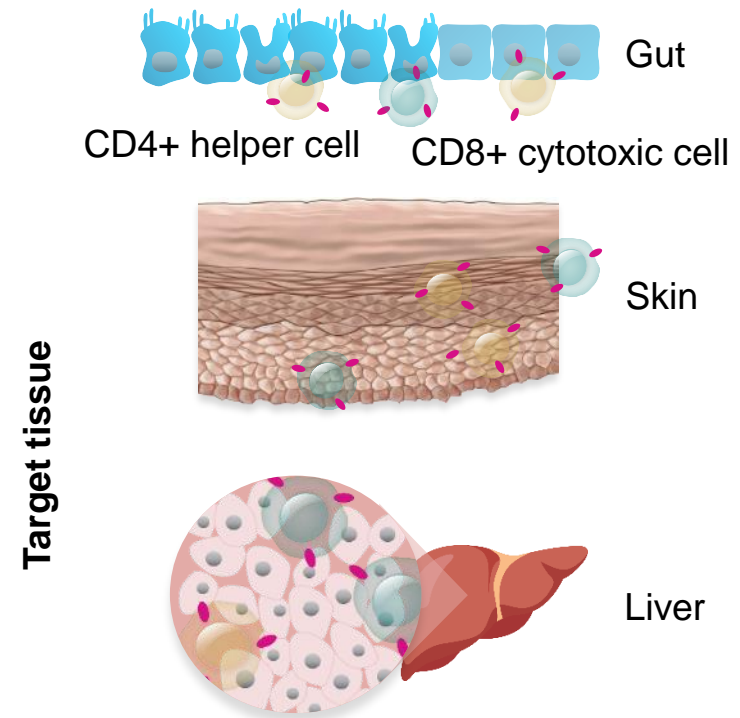
1. Villa NY, et al. *Viruses*. 2016;8:1-30. 2. Abboud R, et al. *Ther Adv Hematol*. 2020;11:1-13. 3. Nassereddine S, et al. *Anticancer Res*. 2017;37:1547-1555. 4. McDonald-Hyman C, et al. *Sci Transl Med*. 2015;7:280-282.

End-Organ Damage Potentiates aGVHD

- Donor T-cell trafficking to target organs is mediated by chemokines, chemokine receptors, and adhesion molecules¹⁻³
- Recruited T cells from lymphoid tissues⁴⁻⁶
 - Proliferate and differentiate into effector T cells
 - Migrate to target organs
 - Release additional cytokines and other immune effectors to cause inflammation, organ damage, and apoptosis
- Sustained end-organ tissue damage potentiates aGVHD through cytokine-mediated feedback loops and maintenance of inflammation²

End-organ damage^{4,7}

Cytolytic donor cells cause end-organ damage and perpetuate pathway propagation



1. Ferrara JLM, et al. *Lancet*. 2009;373:1550-1561. 2. McDonald-Hyman C, et al. *Sci Transl Med*. 2015;7:280-282. 3. Schroeder MA, et al. *Biol Blood Marrow Transplant*. 2018;24:1125-1134. 4. Villa NY, et al. *Viruses*. 2016;8:1-30. 5. Blazar BR, et al. *Nat Rev Immunol*. 2012;12:443-458. 6. Ball LM, et al. *Bone Marrow Transplant*. 2008;41(suppl 2):S58-S64. 7. Nassereddine S, et al. *Anticancer Res*. 2017;37:1547-1555.



Chronic GVHD

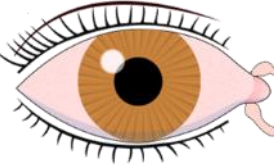
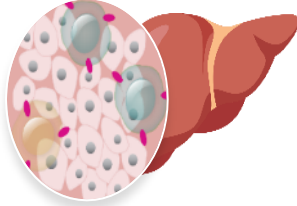
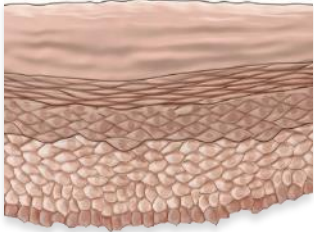

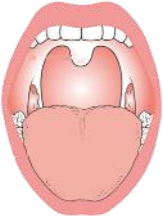
Mechanism of Disease



Clinical Manifestations of cGVHD

- cGVHD is the most common long-term complication of allo-HSCT¹
- Among patients receiving allo-HSCT, 30% to 50% experience cGVHD²⁻⁵
- cGVHD has a median onset of 4 to 6 months after allo-HSCT^{6,7}

Common symptoms⁹

	Eyes	Liver	Skin	Lung	Mouth
					
	cGVHD of the eye can have characteristic hypervascularization ⁸	Inflammatory cellular infiltrates may occur in the bile duct of a patient with cGVHD ⁸	Localized sclerosis of the skin occurs in cGVHD ⁸	Pronounced fibrotic changes can be seen in the lungs of a patient with cGVHD ⁸	Oral cGVHD is characterized by reduced and sclerotic gingiva ⁸
	Dry eyes	Jaundice	Dyspigmentation	Bronchiolitis	Dry mouth

1. Lee SJ. *Blood*. 2017;129:30-37. 2. Kitco CL, et al. *Transplant Cell Ther*. 2021;27(7):545-557. 3. Im A, et al. *Biol Blood Marrow Transplant*. 2020;26(8):1459-1468. 4. Arai S, et al. *Biol Blood Marrow Transplant*. 2015;21(2):266-274. 5. Arora M, et al. *Biol Blood Marrow Transplant*. 2016;22(3):449-455. 6. Lee SJ. *Best Pract Res Clin Haematol*. 2010;23:529-535. 7. Garnett C, et al. *Ther Adv Hematol*. 2013;4:366-378. 8. Zeiser R, Blazer BR. *N Engl J Med*. 2017;377:2565-2579. 9. Ferrara JLM, et al. *Lancet*. 2009;373:1550.

cGVHD: Epidemiology and Significance

cGVHD is associated with significant morbidity¹⁻⁴

- Many patients require ongoing immunosuppressive therapy years after diagnosis of cGVHD
 - A large proportion of patients may need immunosuppressive therapy years after diagnosis depending on severity of disease^{5,6}
- cGVHD is associated with frequent and severe infections^{2,4,7}
- cGVHD leads to debilitating fibrotic organ damage that can be irreversible⁸

cGVHD is associated with worse patient-reported outcomes compared with healthy individuals, including^{1,9-11}

- Significantly lower health-related quality of life
- Decreased functional status
- Inability to work or resume social roles

Among patients who are disease-free after allo-HSCT, cGVHD is a leading cause of NRM^{1,12,13}

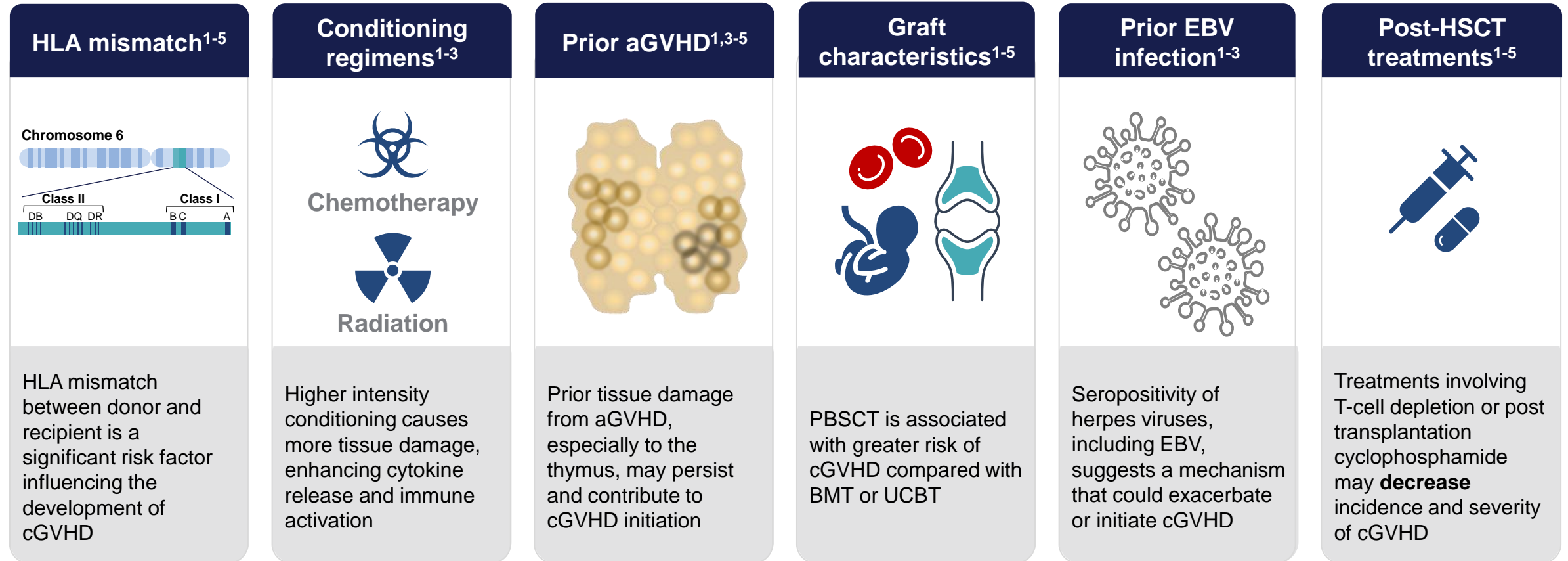
- 60% to 80% 2-year OS and RFS rates^{3,a}
- cGVHD-associated NRM increases over time and is associated with organ failure and infection¹³

^a Two-year survival outcomes are based on data from the Fred Hutchinson Cancer Research Center.³

NRM, non-relapse mortality; OS, overall survival; RFS, relapse-free survival.

1. Lee SJ. *Blood*. 2017;129:30-37. 2. Blazar BR, et al. *Nat Rev Immunol*. 2012;12:443-458. 3. Lee SJ. *Best Pract Res Clin Haematol*. 2010;23:529-535. 4. Garnett C, et al. *Ther Adv Hematol*. 2013;4:366-378. 5. Stewart BL, et al. *Blood*. 2004;104:3501-3506. 6. Curtis LM, et al. *Biol Blood Marrow Transplant*. 2017;23:1980-1988. 7. Socie G, Ritz J. *Blood*. 2014;124:374-384. 8. Hill GR, et al. *Annu Rev Immunol*. 2021 Apr 26;39:19-49. 9. Pidala J, et al. *Blood*. 2011;117:4651-4657. 10. Lee SJ, et al. *Haematologica*. 2018;103:1535-1541. 11. Kurosawa S, et al. *Biol Blood Marrow Transplant*. 2019;25:1851-1858. 12. Wingard JR, et al. *J Clin Oncol*. 2011;29:2230-2239. 13. DeFilipp Z, et al. *Blood Adv*. 2021;5(20):4278-4284.

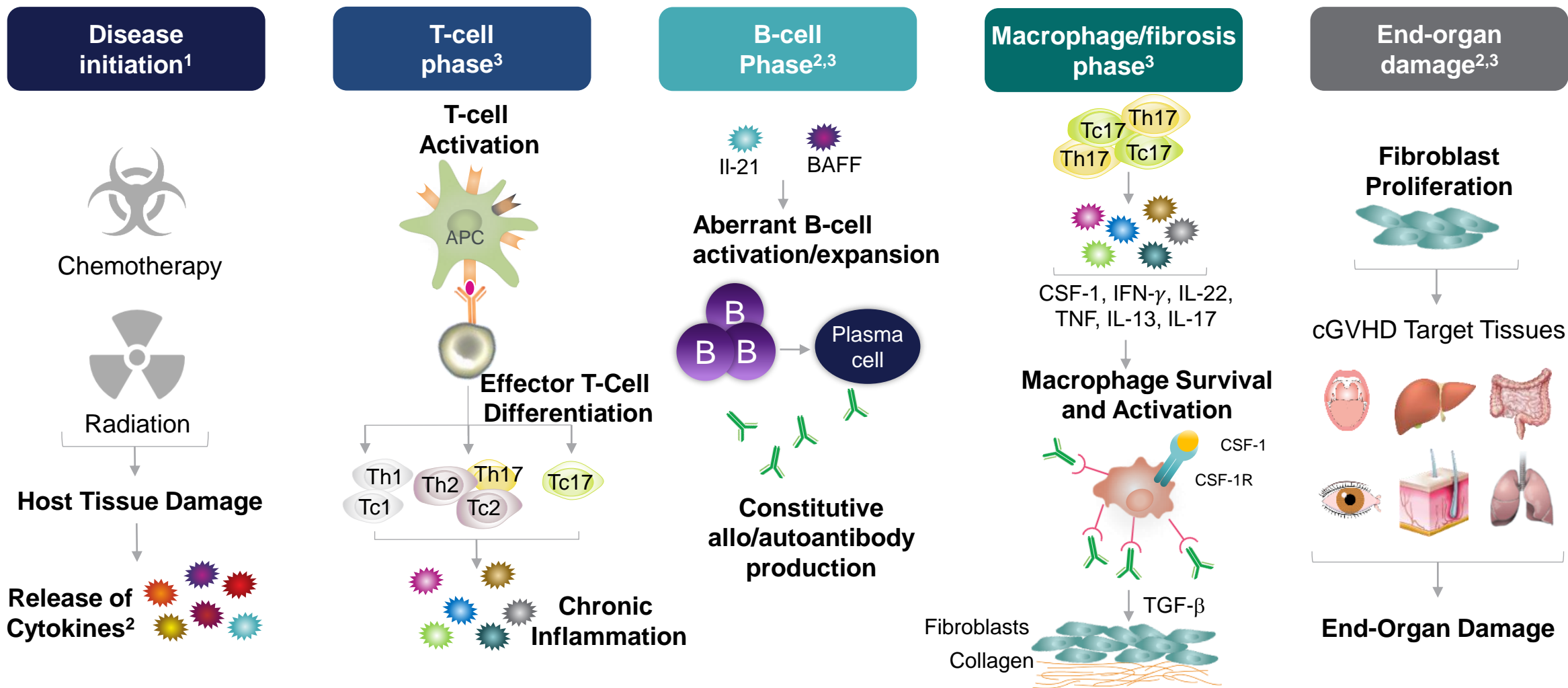
Risk Factors for Development of cGVHD



BMT, bone marrow transplant; EBV, Epstein-Barr virus; PBSCT, peripheral blood stem cell transplant; UCBT, umbilical cord blood transplant.

1. Zeiser R, Blazar BR. *N Engl J Med.* 2017;377:2565-2579. 2. Styczynski J, et al. *J Clin Oncol.* 2016;34:2212-2220. 3. Cooke KR, et al. *Biol Blood Marrow Transplant.* 2017;23:211-234. 4. Lee SJ. *Blood.* 2017;129:30-37. 5. Filipovich AH, et al. *Biol Blood Marrow Transplant.* 2005;11:945-956.

Review of cGVHD Pathophysiology



BAFF, B cell activating factor; CSF, colony-stimulating factor; Tc, cytotoxic T cell; TGF- β , transforming growth factor beta.

1. Toubai T, et al. *Front Immunol.* 2016;7:539. 2. Cooke KR, et al. *Biol Blood Marrow Transplant.* 2017;23:211-234. 3. MacDonald KPA, et al. *Blood.* 2017;129:13-21.

Disease Initiation: Damage to the Host Tissue

1

Disease initiation

1 Response to conditioning regimen-induced tissue damage

Conditioning enables host recipients to receive immunocompetent T cells^{1,2}



Radiation



Chemotherapy

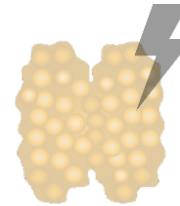


Damage

Conditioning regimens may comprise a combination of chemotherapy, radiation, and/or immunosuppressive drugs

2 Host tissue damage

Damage to the thymus from the conditioning regimen or other causes^a results in impaired negative selection, leading to the development of host-derived donor reactive T cells²⁻⁴



Thymus

Conditioning-mediated damage to host mucosa and skin initiates GVHD⁵



Mouth



Eyes



Liver



Skin



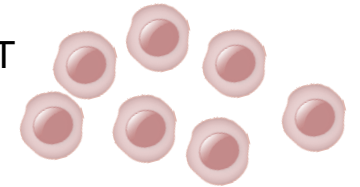
Gut



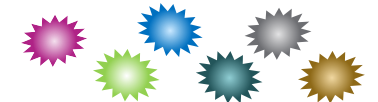
Lungs

3 Cytokine release and acute inflammation

Donor reactive T cells



Release of cytokines



Damage to host tissues causes release of cytokines, leading to^{3,5}

- Donor T-cell differentiation into Th1/Tc1, Th2/Tc2, and Th17 effector cells^{6,7}
- Recruitment of donor T cells into the target tissue^{6,7}

^aOther causes of thymic injury include prophylaxis with calcineurin inhibitors, alloreactive T cells, low serum thymic hormone levels, and immunoglobulin deposition.³

1. Ferrara JL, et al. *Stem Cells*.1996;14:473-489. 2. Blazar BR, et al. *Nat Rev Immunol*. 2012;12:443-458. 3. Zeiser R, Blazar BR. *N Engl J Med*. 2017;377:2565-2579.

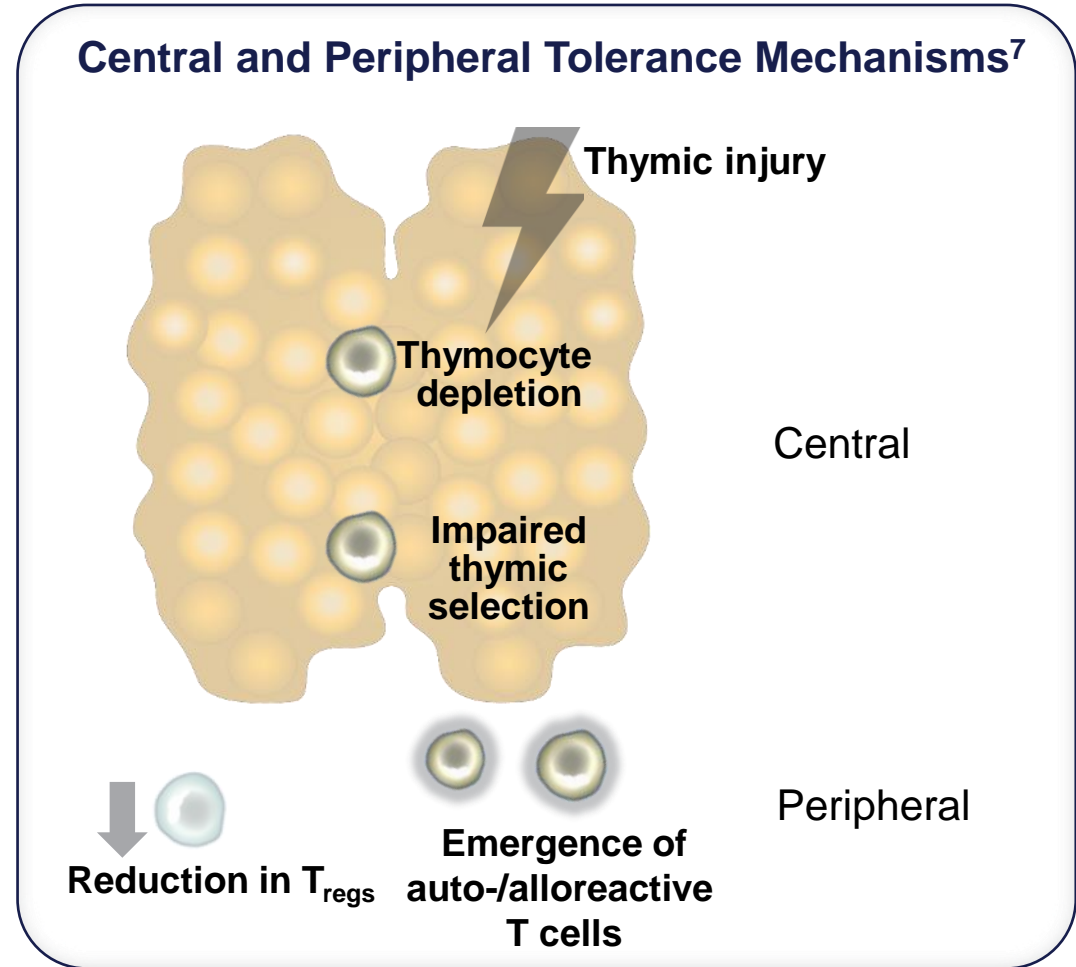
4. Soares MV, et al. *Front Immunol*. 2019;10:334. 5. Toubai T, et al. *Front Immunol*. 2016;7:539. 6. Cooke KR, et al. *Biol Blood Marrow Transplant*. 2017;23:211-234. 7. Schroeder MA, DiPersio JF. *Dis Model Mech*. 2011;4:318-333.

Dysregulated T-Cell Immunity: Thymic Dysfunction Contributes to cGVHD

2

T-cell phase

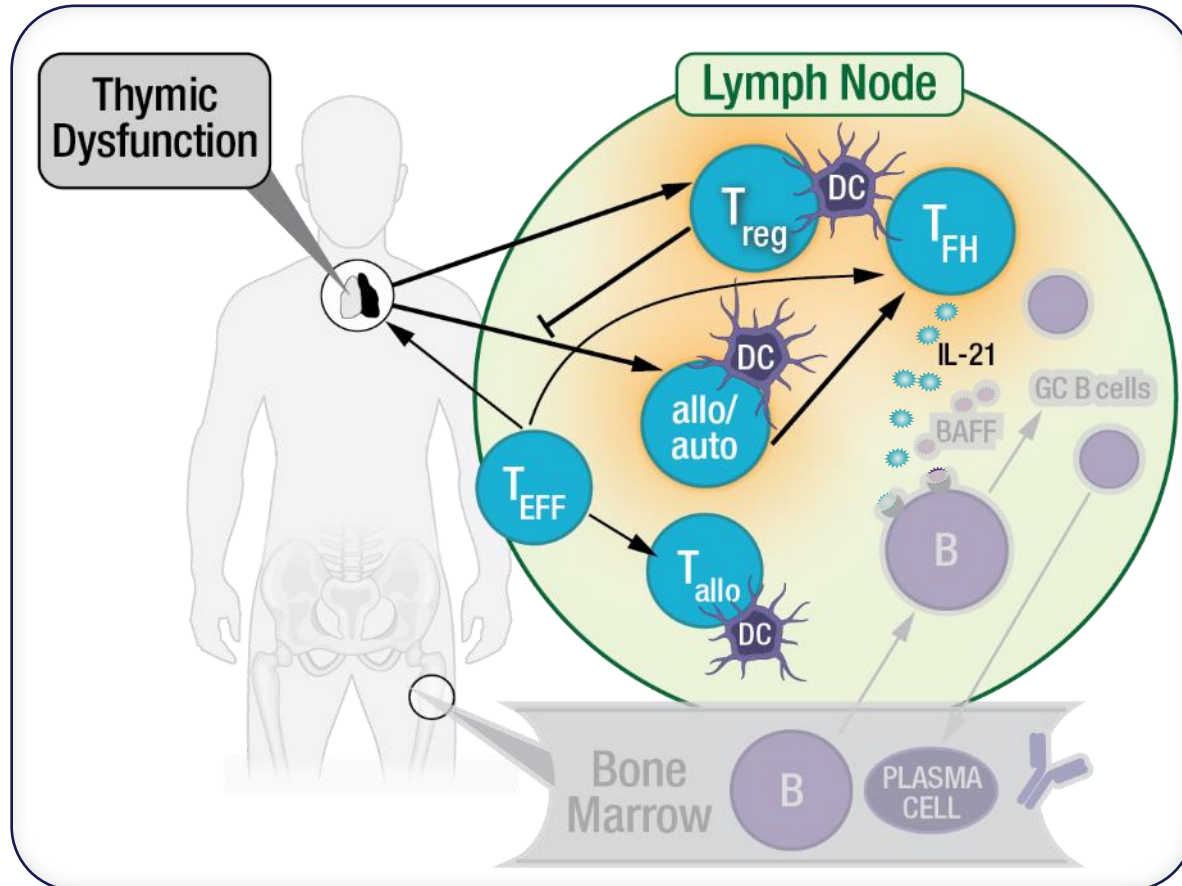
- Thymic injury, caused by the conditioning regimen, alloreactive T cells, or prior aGVHD, leads to¹⁻⁶
 - Emergence of alloreactive and autoreactive T cells due to **impaired thymic selection**
 - **Thymocyte depletion** resulting in the loss of T_{regs} , further facilitating escape of auto- and alloreactive T cells into the periphery
- Disrupted immune regulation leads to activation of autoreactive and alloreactive T cells, further propagating cGVHD pathology^{2,5-7}



1. Cooke KR, et al. *Biol Blood Marrow Transplant*. 2017;23:211-234. 2. MacDonald KPA, et al. *Blood*. 2017;129:13-21. 3. Wu T, et al. *J Immunol*. 2013;191:488-499. 4. Zeiser R, Blazar BR. *N Engl J Med*. 2017;377:2565-2579. 5. McDonald-Hymen C, et al. *Sci Transl Med*. 2015;7:280-282. 6. Beres AJ, Drobyski WR. *Front Immunol*. 2013;4:1-9. 7. Gregersen PK, Behrens TW. *Nat Rev Genet*. 2006;7:917-928.

Dysregulated T-Cell Immunity: An Imbalance Between Effector Cells and T_{regs}

T-cell phase



- Dysregulation of immune mechanisms via¹⁻⁵
 - Donor T-cell activation
 - Autoreactive T-cell proliferation
 - Reduced T_{regs}
- Activation of donor lymphocytes leads to differentiation into effector populations^{1,3,5}
- Cytolytic attack by effector T cells leads to further recruitment of immune cells¹
- Proinflammatory cytokines and an imbalance in effector and regulatory populations promote chronic inflammation, leading to widespread tissue fibrosis and exacerbating cGVHD¹⁻⁷

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DC, dendritic cell; T_{EFF}, effector T cell; T_{FH}, T follicular helper cell.

1. Cooke KR, et al. *Biol Blood Marrow Transplant*. 2017;23:211-234. 2. Blazar BR, et al. *Nat Rev Immunol*. 2012;12:443-458. 3. Zeiser R, Blazar BR. *N Engl J Med*. 2017;377:2565-2579.

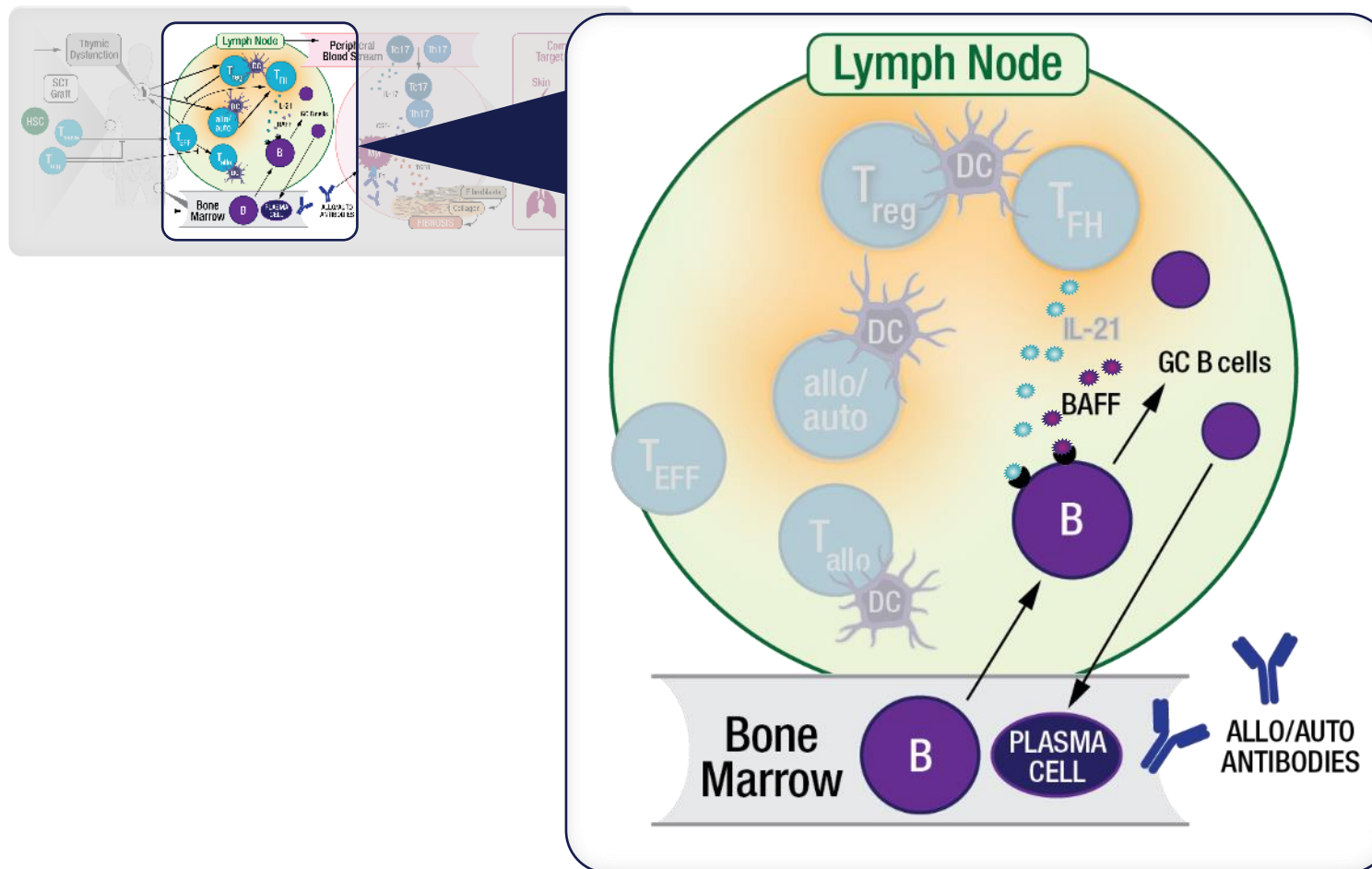
4. MacDonald KPA, et al. *Blood*. 2017;129:13-21. 5. Schroeder MA, DiPersio JF. *Dis Model Mech*. 2011;4:318-333. 6. MacDonald KPA, et al. *J Clin Invest*. 2017;127:2452-2463.

7. McDonald-Hymen C, et al. *Sci Transl Med*. 2015;7:280-282.

Aberrant B-Cell Activation and Expansion

3

B-cell phase



- Aberrant B-cell activation and expansion occurs due to
 - Impaired B-cell reconstitution after HSCT¹⁻³
 - Elevated expression of BAFF^{1,3}
 - Interactions with donor T cells^{3,4}
- Activated B cells capable of constitutive **allo/autoantibody production** contribute to the inflammatory response and target tissue damage¹⁻⁷

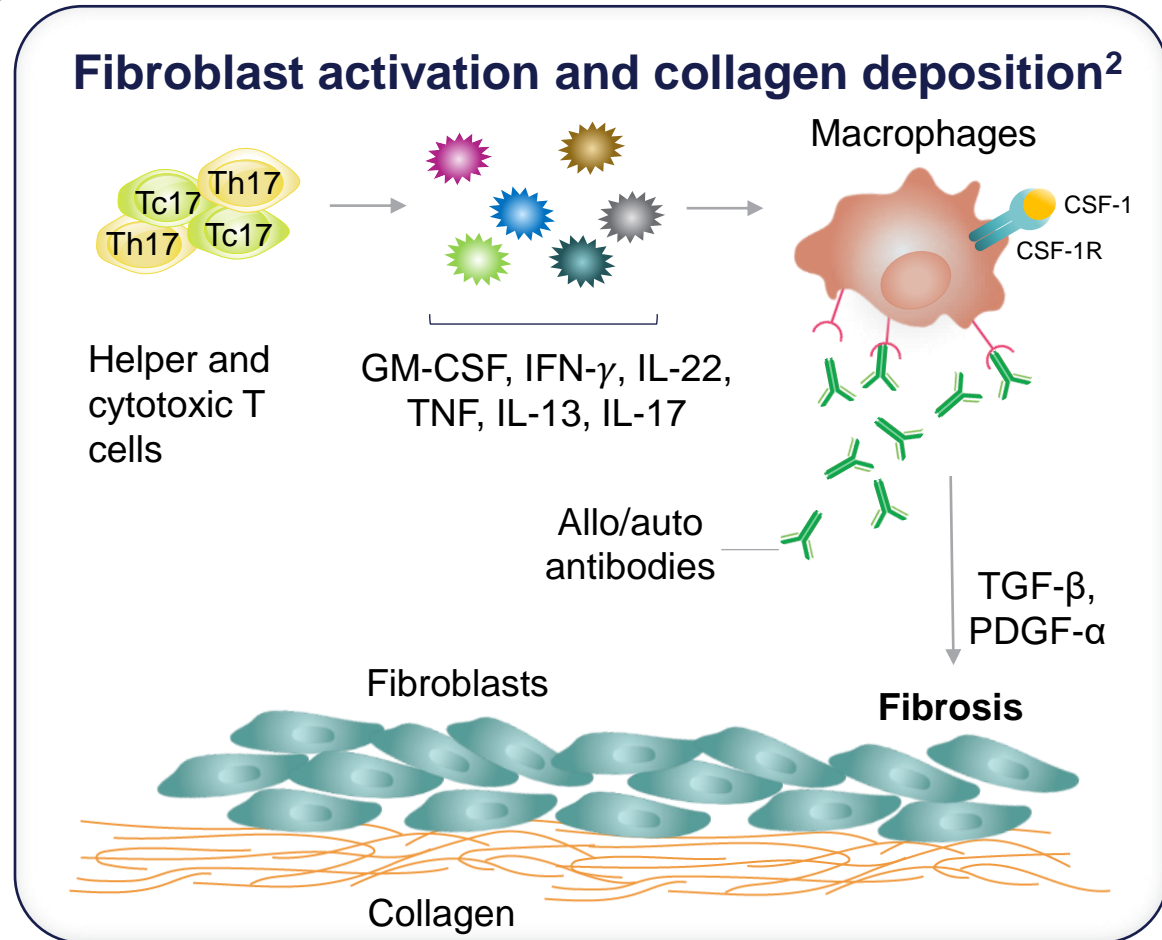
Reproduced from *Blood*, Vol 129, MacDonald KPA, et al. Chronic graft-versus-host disease: biological insights from preclinical and clinical studies, Pages 13-21, Copyright (2017), with permission from Elsevier.
GC, germinal center.

1. Sarantopoulos S, et al. *Biol Blood Marrow Transplant*. 2015;21:16-23. 2. Shimabukuro-Vornhagen A, et al. *Blood*. 2009;114:4919-4927. 3. Cooke KR, et al. *Biol Blood Marrow Transplant*. 2017;23:211-234. 4. Schroeder MA, DiPersio JF. *Dis Model Mech*. 2011;4:318-333. 5. MacDonald KPA, et al. *Blood*. 2017;129:13-21. 6. Zeiser R, Blazar BR. *N Engl J Med*. 2017;377:2565-2579. 7. McDonald-Hyman C, et al. *Sci Transl Med*. 2015;7:280rv2.



Activated Macrophages Produce Inflammatory Cytokines and Growth Factors That Drive Fibrosis

- Effector cell–produced proinflammatory cytokines and B-cell–derived allo/ autoantibodies promote macrophage activation¹⁻⁴
- Donor-derived macrophages are dependent on CSF-1R signaling for proliferation, differentiation, and migration⁵⁻¹¹
- CSF-1R–dependent activated macrophages mediate production of TGF- β , which contributes to intestinal pathology, epidermal inflammation, and subcutaneous and cutaneous fibrosis in cGVHD¹²⁻¹³



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GM-CSF, granulocyte-macrophage colony-stimulating factor; PDGF- α , platelet-derived growth factor alpha.

1. MacDonald KPA, et al. *Blood*. 2017;129:13-21. 2. MacDonald KPA, et al. *J Clin Invest*. 2017;127:2452-2463. 3. Cooke KR, et al. *Biol Blood Marrow Transplant*. 2017;23:211-234. 4. Schroeder MA, DiPersio JF. *Dis Model Mech*. 2011;4:318-333. 5. Hume DA, et al. *J Immunol*. 1988;141(10):3405-3409. 6. Hume DA and MacDonald KP. *Blood*. 2012;119(8):1810-1820. 7. Kelley TW, et al. *J Biol Chem*. 1999;274(37):26393-26398. 8. Mossadegh-Keller N, et al. *Nature*. 2013;497(7448):239-243. 9. Rieger MA, et al. *Science*. 2009;325(5937):217-218. 10. Stanley ER and Chitu V. *Cold Spring Harb Perspect Biol*. 2014;6(6):a021857. 11. Tushinski RJ and Stanley ER. *J Cell Physiol*. 1983;116(1):67-75. 12. Alexander KA, et al. *J Clin Invest*. 2014;124(10):4266-4280. 13. Banovic T, et al. *Blood*. 2005;106(6):2206-2214.

Aberrant Tissue Repair, Propagation of Fibrosis, and Progression of cGVHD¹

End-organ damage

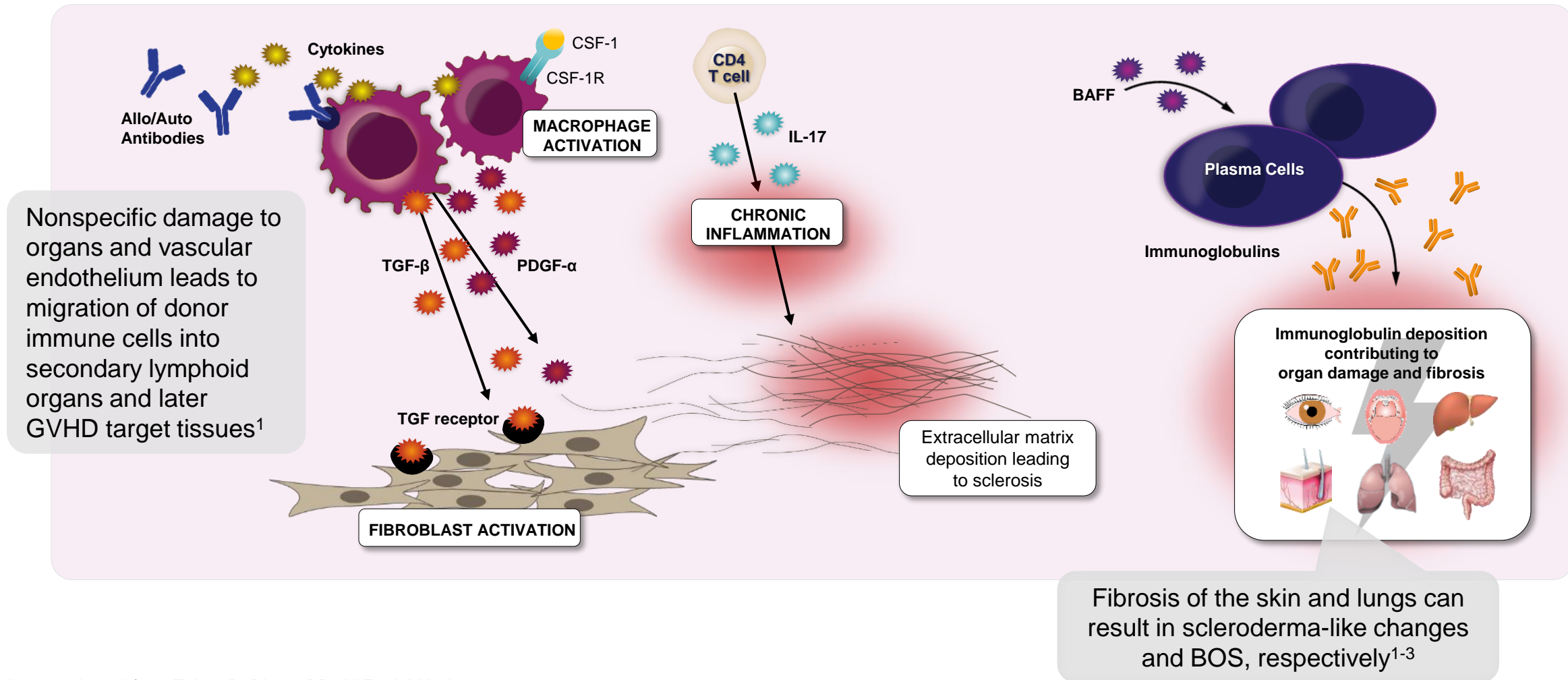


Image adapted from Zeiser R, Blazar BR. *N Engl J Med.* 2017;377:2565-2579.
 BOS, bronchiolitis obliterans syndrome.

1. Cooke KR, et al. *Biol Blood Marrow Transplant.* 2017;23:211-234. 2. MacDonald KPA, et al. *J Clin Oncol.* 2017;127:2452-2463. 3. Kitko CL, et al. *Biol Blood Marrow Transplant.* 2012;18:S46-S52.



Appendix



Histologic Features of cGVHD

- Displays autoimmune and fibrotic features¹
- Inflammatory proteins and fibrotic changes cause diffuse, nonspecific damage to numerous organs and the vascular endothelium^{1,2}

End Organ/Site	Characteristic Histopathologic Findings ³
Liver	<ul style="list-style-type: none"> • Immune-mediated damage to small bile ducts and ductules • Cholestatic and inflammatory changes
Skin	<ul style="list-style-type: none"> • Superficial interface dermatitis with vacuolar change in the basilar layer • Lichenoid pattern of lymphocytic inflammation ± lymphocyte satellitosis
GI	<ul style="list-style-type: none"> • Destruction of basilar glands or crypts • Mucosal denudation • Enterocyte apoptosis
Mucosa (eg, oral cavity, eye)	<ul style="list-style-type: none"> • Exocytosis, apoptosis, and/or lichenoid interface inflammation characteristic of localized or generalized epithelial changes • Conjunctival features include lymphocyte exocytosis, satellitosis, vacuolization of the basal epithelium, and epithelial cell necrosis
Lungs	<ul style="list-style-type: none"> • Constrictive bronchiolitis obliterans (cGVHD) • Cryptogenic organizing pneumonia (aGVHD and cGVHD)

Note: Diagnosis of GVHD is not achieved by histopathology alone; histology results must be integrated into the context of clinical presentation.²

GI, gastrointestinal.

1. Blazar BR, et al. *Nat Rev Immunol*. 2012;12:443-458. 2. Cooke KR, et al. *Biol Blood Marrow Transplant*. 2017;23:211-234. 3. Shulman HM, et al. *Biol Blood Marrow Transplant*. 2015;21:589-603.

Potential Biomarkers for cGVHD

Biomarker	Biologic Function	Prognostic	Diagnostic	Predictive
BAFF	BAFF is a survival factor for B cells and controls normal B-cell maturation; however, it promotes survival of autoreactive B cells ^{1,2} ; increased levels of sBAFF correlate with onset of, and active cGVHD. Increased levels 1 month after ECP predicted response of skin cGVHD ^{3,4}		✓	✓
B cells	A breakdown in peripheral B-cell tolerance and altered B-cell homeostasis are components of cGVHD ¹ ; an imbalance of certain B-cell subsets is associated with the diagnosis and severity of cGVHD ^{4,5}	✓	✓	✓
CD163	CD163 is a macrophage scavenger receptor regulated by inflammatory mediators ⁶ ; plasma CD163 associates with de novo-onset cGVHD ^{3,4}	✓		
CTLA-4 SNP	Association between position +49 guanine to guanine homozygote genotype in donors and higher risk of cGVHD ^{5,7}	✓		
CXCL9	T cell type 1 chemokine detected in the blood that attracts CXCR3+ T cells in cGVHD target organs ⁸ ; increased levels and up-regulated gene expression detected at diagnosis of cGVHD; increased levels at symptom onset associated with severe cGVHD ³⁻⁵	✓	✓	
CXCL10	Increased levels and up-regulated gene expression detected at diagnosis and in active cGVHD ³⁻⁵	✓	✓	
DKK3	Glycoprotein that regulates Wnt signaling, RYK and Ror ⁸ ; increased levels at diagnosis associated with NRM ^{3,4}	✓	✓	
IL-15	Homeostatic cytokine ⁵ ; low levels of IL-15 on day 7 after allo-HSCT associated with a 2.7-fold higher likelihood of developing cGVHD ^{4,9}	✓	✓	
IL-2Rα	Decrease in sIL-2R α predicts response to the combination of ECP and ruxolitinib ⁵			✓
miRNAs	Play a regulatory role in immune response and autoimmunity; miR-155 strongly associated with onset and severity of cGVHD, and differentially expressed in patients with severe and mild cGVHD ⁵		✓	

CTLA, cytotoxic T-lymphocyte antigen; CXCL, C-X-C Motif Chemokine Ligand; DKK, dickkopf; ECP, extracorporeal photophoresis; miRNAs, micro RNAs, NRM, nonrelapse mortality; Ror, receptor tyrosine kinase-like orphan receptor; RYK, receptor-like tyrosine kinase; sBAFF, soluble B-cell activating factor; SNP, single-nucleotide polymorphism; TRM, treatment-related mortality.

1. Sarantopoulos S, et al. *Blood*. 2009;113:3865-3874. 2. Liu Z, Davidson A. *Trends Immunol*. 2011;32:388-394. 3. Bidgoli A, et al. *Transpl Cel Ther*. 2022;28:657-666. 4. Milosevic E, et al. *Front Immunol*. 2022;13:1033263. doi: 10.3389/fimmu.2022.1033263. 5. Ji R, et al. *Crit Rev Oncol / Hematol*. 2023;186:103993. <https://doi.org/10.1016/j.critrevonc.2023.103993>. 6. Inamoto Y, et al. *Biol Blood Marrow Transplant*. 2017;23:1250-1256. 7. Wang Z, et al. *Hematol*. 2021;26(1):144-153. 8. Logan BR, et al. *J Clin Invest*. 2023;133(15):e168575. 9. Pratt LM, et al. *Bone Marr Tranpl*. 2013;48(5):722-728.

Potential Biomarkers for cGVHD (cont)

Biomarker	Biologic Function	Prognostic	Diagnostic	Predictive
MMP3	MMP family of proteins regulates the breakdown of extracellular matrix and tissue remodeling ¹ ; increased MMP3 levels in BOS patients ²		✓	
MMP9	MMP family of proteins regulates the breakdown of extracellular matrix and tissue remodeling ¹ ; Increased levels at BOS diagnosis associated with OS ²	✓		
NK/NK_{reg} cells	Play a key role in tissue fibrosis ³ ; along with an imbalance of other immune cell compartments, loss of NK _{reg} cells associates with the future development and severity of cGVHD ⁴ and different subpopulations may have potential in diagnosing organ-specific cGVHD ³	✓	✓	
Osteopontin	Osteopontin has broad biological functions, including biomineralization, bone remodeling, and inflammation ⁵ ; detection in plasma as part of a panel with ST2, CXCL9 and MMP3 can predict future cGVHD ⁶	✓	✓	
anti-PDGFR	Levels correlate with cGVHD diagnosis and severity; predictive of response to nilotinib for steroid-refractory or steroid-dependent cGVHD ^{3,7}	✓	✓	✓
PD-1	Inhibitory immune checkpoint receptor; maintains immune tolerance. ³ Levels of soluble PD-1 decreased in cGVHD patients; related to OS, DFS and TRM, and PD-1 expression in circulating CD4+ and CD8+ T cells may be predictive of response to abatacept ^{3,8}	✓	✓	
Reg3α	Peptide primarily found in Paneth cells of the intestines; increased levels associated with GI cGVHD; increased levels at GI cGVHD diagnosis associated with NRM ²	✓	✓	
anti-Ro52	Autoantibody against the Ro52 protein; levels found to be higher in patients with active cGVHD than without active cGVHD, and associated with severity of cGVHD; diagnostic potential for skin and liver cGVHD ^{3,9}	✓		
ST2	Interaction between ST2 and its ligand, IL-33, triggers inflammatory cytokine production and cell proliferation ¹⁰ ; ST2 levels declined after 2, 4, and 6-months of ECP ²			✓
Th17 cells	Increased plasma Th17 cells at cGVHD onset; higher levels in allografts predicts high treatment sensitivity ³		✓	✓
T_{reg} cells	T _{regs} establish tolerance between recipient tissues and donor-derived immunity; decreased T _{reg} in peripheral blood in cGVHD, but enriched in target tissues. ³ Predicts response to ECP in steroid-refractory cGVHD ³		✓	✓

DFS, disease-free survival; MMP, matrix metalloproteinase; NK_{reg}, regulatory natural killer cell; OS, overall survival; PD-1, programmed cell death protein-1; Reg3α, regenerating islet-derived 3α; ST2, suppression of tumorigenicity 2; TRM, treatment-related mortality.

1. Wang X, Khalil RA. *Adv Pharmacol.* 2018;81:241-330. 2. Bidgoli A, et al. *Transpl Cel Ther.* 2022;28:657-666. 3. Ji R, et al. *Crit Rev Oncol / Hematol.* 2023;186:103993. <https://doi.org/10.1016/j.critrevonc.2023.103993>. 4. Schultz KR, et al. *Blood.* 2020;135:1287-1298. 5. Icer MA, Gezmen-Karadag M. *Clin Biochem.* 2018;59:17-24. 6. Logan BR, et al. *J Clin Invest.* 2023;133(15):e168575. 7. Chen GL, et al. *Biol Blood Marr Transpl.* 2018;24(2):373-380. 8. Kordelas L, et al. *Bone Marr Transpl.* 2021; 12:3799. 9. Yang K, et al. *Front Immunol.* 2020;11:1505. 10. Reichenbach DK, et al. *Blood.* 2015;125:3183-3192.



